Animal Research Protocols Involving Hazardous Chemicals

I. OVERVIEW

Hazardous Chemicals: Known or suspect carcinogens, reproductive toxins or other highly toxic substances (e.g., anti-neoplastic agents), and nanomaterials may be used in laboratory animal research protocols. Reference the SU Chemical Hygiene Plan for hazardous chemical definitions.

Potential Exposures: Research staff may be exposed to these hazardous chemicals during preparation, handling, and animal dosing. These substances may be excreted and/or secreted from the animal and, therefore, be present in the animal's bedding in extremely low concentrations. Animal husbandry staff and veterinarians may be exposed to these hazardous chemicals or their toxic metabolites during cage handling or when handling medicated water or feed.

How to Use this Guide: This guidance provides information for Principal Investigators to consider when planning and conducting their animal protocol involving hazardous chemicals so that appropriate controls are established in order to minimize potential exposure of laboratory personnel, husbandry staff, and veterinarians. See Section IV for a quick guide summarizing the general safety practices for: hazardous chemical preparation and handling; animal dosing; and cage management.

II. PLANNING YOUR ANIMAL PROTOCOL

1. Identify Hazards of Chemicals

SU Chemical Hygiene Plan
The use of hazardous chemicals in animal research protocols falls under the SU Chemical Hygiene Plan.

As with any laboratory operation, the Principal Investigator (PI) and researchers must identify and understand the hazards associated with the chemical (e.g., toxicity, reactivity, flammability, corrosivity, etc.) before they begin their work so that appropriate controls can be established.

Intrinsic Hazards of Chemical
Review Material Safety Data Sheets (MSDSs) and other sources of safety information to understand the hazards of the chemical and any other special considerations.

Suggested Sources of Safety Information on Hazardous Chemicals

- SU's MSDS database: [http://msds.stanford.edu](http://msds.stanford.edu)
- SU's Laboratory Chemical Safety Toolkit chemical hazard classes:
Animal Metabolism of the Chemical
It is important to understand how the chemical is metabolized in a specific animal species so that appropriate controls can be implemented for safe animal and cage management after dosing. A review of peer-reviewed literature may yield this information, but it is often not documented. In the absence of such data, follow controls provided in Section III below.

Medical Surveillance
Medical surveillance and/or use of an antidote may, on the rare occasion, be required if the hazardous chemical used in an animal protocol is known to pose a serious health risk, such as diphtheria toxin or 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). The PI must identify these chemicals during the safety literature review for the experiment and consult with EH&S’s Occupational Health & Safety Program at 723-0448.

Submit Protocol for APLAC Review

- Complete the Administrative Panel on Laboratory Animal Care (APLAC) eProtocol form.
  - Describe the proposed use of hazardous chemicals in question #8 of the “Are You Using?” section.
  - Indicate that controls provided in Section III below will be implemented or its equivalent as described in your lab’s Standard Operating Procedure (SOP).

Failure to properly identify hazardous chemical use may result in inadequate provision of chemical safety measures and may delay APLAC approval.

Coordinate with the Veterinary Service Center

- Contact the Veterinary Service Center (VSC) at 723-3876 at least two weeks before beginning the project to obtain:
  - Permission to use the facility
  - Room assignment
  - Provisions for appropriate cage labeling and waste management

- Provide a copy of the MSDS or other hazard information so the VSC can ensure VSC personnel have chemical-specific hazard training.
III. CONDUCTING YOUR ANIMAL PROTOCOL

1. Hazardous Chemical Preparation and Handling

- Follow the Standard Operating Procedure (SOP) established by the Principal Investigator. Guidance on SOP prioritization and development is available at SU’s Laboratory Chemical Safety Toolkit. At minimum, handle hazardous chemicals using:
  - Laboratory fume hood or other appropriate engineering control.
  - Appropriate personal protective equipment (PPE): safety eyewear, chemical-resistant gloves appropriate for the chemical, and lab coat over appropriate street clothing (long pants and closed-toe shoes).

- Ensure research personnel are trained on the SOPs and specific hazards associated with the chemicals. Ensure proper waste management. Maintain training documentation for at least one year.

2. Animal Dosing and Care

- Wear appropriate personal protective equipment: disposable solid-front gown, head and shoe coverings, safety glasses/goggles, and chemical-resistant gloves (double gloving recommended).

- Appropriately restrain or sedate animals per your APLAC protocol to reduce the possibility of accidental self-inoculation. Contact the VSC if your procedure does not allow for restraints or sedation.

- If administering hazardous chemical by:
  - **Injection**: Use safety syringe and follow sharps handling procedures as described in the Biosafety Manual. If unable to use a safety syringe, you must document the reasons for this.
  - **Tablet**: Avoid liberating dust.
  - **Topical Application**: Avoid direct dermal contact with animal's application site.
  - **Water/Feed**: Minimize aerosolization (e.g., do not shake feed into dispenser).
  - **Aerosolization**: Use the fume hood or biosafety cabinet ducted to the outdoors. If unable to use these equipment, contact EH&S for a respirator assessment.

- Place animals back into cage and label cage with “Pink Card” (see below).

- Decontaminate lab surfaces and equipment using wet wiping methods with appropriate cleaning agent.

- Dispose sharps, chemical waste and PPE properly. Wash hands after removing gloves.
Cage Management

HOUSING

- If feasible, consider using disposable cages or alternative bedding (e.g., paper liner) to minimize potential aerosolization of the hazardous chemical during cage changing.
- Maintain cages on ventilated racks or with microisolator lids.

SIGNAGE

- Label cages with the “Pink Card” as follows:
  - Indicate the following information: “Animals treated with [name of chemical]. Excreta/bedding may be contaminated.” Affix “Hazardous Drug” sticker to card.
  - Indicate dosage and date/time animal was dosed.
  - Also label water bottle/feeder if hazardous chemical is administered via water/feed.
- Maintain label on cage for 72 hours after last dosing AND until contaminated bedding is changed, unless longer time frames are required as identified in the risk assessment during the planning phase.

CAGE CLEANING/BEDDING MANAGEMENT

*For all cage cleanings/bedding disposals performed up to 72 hours after animal dosing AND until contaminated bedding is changed, unless risk assessment requires longer time frame.*

- Wear appropriate personal protective equipment (PPE): long-sleeved lab coat or disposable gown, plastic apron, rubber boots or disposable plastic booties, chemical-resistant gloves, and safety glasses. Also use goggles/face shield where splash potential exists.
- Place contaminated cage (with bedding, water and feed) into the bin specially marked “Chemotherapy Waste – For Incineration Only” at the ventilated dump station to ensure proper disposal by the VSC. Outside the VSC, a laboratory fume hood should be used to dump contaminated bedding (thoroughly wet down bedding to help minimize dust generation) into a bag and collected in a bin specially marked “Chemotherapy Waste – For Incineration Only” or tagged as chemical waste. To obviate this disposal requirement, contact the EH&S Environmental Programs Manager at 723-0448 for an assessment.
  - Respirators may be required if any of the aforementioned engineering controls are not feasible. Contact EH&S at 723-0448 for guidance.
  - Do not use laminar flow hoods which are commonly found in VSC animal rooms. These units do not provide personnel protection.
- For large animal pens (e.g., pig/dog runs) follow VSC procedures: hose down waste into the sewer, do not direct jet stream at excreta, etc.
- Manage animal carcasses as pathological waste (i.e., tissues must be incinerated).
- Remove gloves and wash hands after working with animals, cages, and contaminated bedding.
CLEANUP OF WATER/FOOD SPILLS
(If hazardous chemical has been added to water/feed)

Assuming that the person is knowledgeable about the hazards of the chemical and has the appropriate spill cleanup materials, proceed as follows for small spills:

- Wear appropriate personal protective equipment (PPE): long-sleeved lab coat, plastic apron, rubber boots or disposable plastic booties, chemical-resistant gloves, and safety glasses. Also use goggles/face shield where splash potential exists.
- Transfer animals to clean cages via VSC procedures.
- At a ducted ventilated dump station or laboratory fume hood, dump contaminated material in the bin specially marked “Chemotherapy Waste – For Incineration Only” to ensure proper disposal.
  - Respirators may be required if any of the aforementioned engineering controls are not feasible. Contact EH&S at 723-0448 for guidance.
  - Do not use laminar flow hoods which are commonly found in VSC animal rooms. These units do not provide personnel protection.
- If unable to or not prepared to clean up a spill, call EH&S at 725-9999 on campus and 286 in the School of Medicine.
IV. SUMMARY OF HAZARDOUS CHEMICAL EXPOSURE CONTROLS:

<table>
<thead>
<tr>
<th>Hazardous Chemical Prep and Handling</th>
<th>Animal Dosing</th>
<th>Cage Management</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Personal Protective Equipment</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>• Safety goggles/glasses</td>
<td>• Disposable solid-front gown with lab coat</td>
<td>• Long sleeved lab coat or gown</td>
</tr>
<tr>
<td>• Chemical-resistant gloves</td>
<td>• Head and shoe coverings</td>
<td>• Plastic apron</td>
</tr>
<tr>
<td>• Long sleeved lab coat</td>
<td>• Safety goggles/glasses</td>
<td>• Rubber boots or disposable plastic booties</td>
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<tr>
<td>• Appropriate street clothing</td>
<td>• Chemical-resistant gloves (double-gloving)</td>
<td>• Chemical-resistant gloves</td>
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<tr>
<td>(long pants and closed-toe shoes)</td>
<td>• Appropriate street clothing</td>
<td>• Safety glasses/goggles (also use face shield when splash potential exists; e.g. hosing down large animal cages, pens and runs)</td>
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<tr>
<td></td>
<td></td>
<td>Note: If engineering controls described below are not feasible, contact EH&amp;S for respirator assessment.</td>
</tr>
<tr>
<td><strong>Work Practices/ Engineering Controls</strong></td>
<td><strong>Labeling/ signage</strong></td>
<td><strong>Disinfection</strong></td>
</tr>
<tr>
<td>Prepare and handle hazardous chemical in a laboratory fume hood, including weighing, mixing, filtering and heating.</td>
<td>• Ensure proper storage and labeling of hazardous chemical.</td>
<td>Decontaminated lab surfaces and equipment with appropriate cleaning agents.</td>
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<tr>
<td>NOTE: Do not use positive laminar flow hoods (commonly found in VSC animals rooms) - air flows towards worker.</td>
<td>• Consult chemical manufacturer’s instructions and/or SU’s Chemical Safety Database.</td>
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<td>Ensure fume hood and eyewash/safety shower are currently certified and maintained.</td>
<td>For injections: Ensure animals are appropriately restrained per your research protocol to reduce possibility of accidental self-inoculation and use safety syringe. Follow sharps handling procedures per Biosafety Manual.</td>
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<tr>
<td>Ensure personnel is trained on the SOP and the specific chemical hazards.</td>
<td>Use fume hood when administering hazardous chemicals by aerosolization. If tablets are administered, avoid liberating dust. Consult EH&amp;S for respirator assessment if fume hood is not used.</td>
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<tr>
<td>Ensure proper disposal of sharps, PPE and chemical waste.</td>
<td>For topical application, avoid dermal contact with animal’s application site.</td>
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<td>Wash hands after removing gloves.</td>
<td>Ensure proper disposal of sharps, PPE and chemical waste.</td>
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<tr>
<td>For injections:</td>
<td>Wash hands after removing chemical-resistant gloves.</td>
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<tr>
<td>Maintain cages on ventilated racks or with microisolator lids.</td>
<td>If hazardous chemical will be present in cage:</td>
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<tr>
<td>Follow VSC guidelines on transporting animals.</td>
<td>• Label the cage with “Pink card” and Hazardous Drug sticker with the following information:</td>
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</tr>
<tr>
<td>Consider using disposable cages or using alternative bedding (e.g., paper liner) to minimize aerosolization of bedding containing hazardous chemicals.</td>
<td>Animals treated with “name of chemical.” Excreta/ bedding/ water/ feed may be contaminated. Do not change cage without wearing appropriate personal protective equipment per the SOP.</td>
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<tr>
<td>Cage cleaning: Dump cages with hazardous chemicals contained in the bedding, food and/or water at the ventilated dump station into collection bin labeled “Chemotherapy waste – For incineration only” or in the fume hood, thoroughly wet down bedding to help minimize dust generation; gently dump into bag and place in collection bin labeled “Chemotherapy waste – For incineration only.”</td>
<td>• Indicate date and time animal was dosed.</td>
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<tr>
<td>For large animal pens (e.g., pig/dog runs) follow VSC cleaning protocols (e.g. hose down sewer, do not direct jet stream at excreta).</td>
<td>• Maintain pink card for 72-hours after last dosing and until contaminated bedding/water/feed is changed.</td>
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<td>Wash hands after removing chemical-resistant gloves.</td>
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